

# Optimization of total flavonoid compound extraction from *Camellia sinensis* using the artificial neural network and response surface methodology

Ivan M. Savić<sup>1</sup>, Vesna D. Nikolić<sup>1</sup>, Ivana M. Savić<sup>1</sup>, Ljubiša B. Nikolić<sup>1</sup>, Mihajlo Z. Stanković<sup>1</sup>, Karl Moder<sup>2</sup>

<sup>1</sup>University of Niš, Faculty of Technology, Leskovac, Serbia

<sup>2</sup>University of Natural Resources and Life Sciences, Institute of Applied Statistics and Computing, Center of Experimental Design, Vienna, Austria

## Abstract

The aim of this paper was to model and optimize the process of total flavonoid extraction from green tea using the artificial neural network and response surface methodology, as well as the comparison of these optimization techniques. The extraction time, ethanol concentration and solid-to-liquid ratio were identified as the independent variables, while the yield of total flavonoid was selected as the dependent variable. Central composite design (CCD), using a second-order polynomial model and multilayer perceptron (MLP) were used for fitting the obtained experimental data. The values of root mean square error, cross-validated correlation coefficient and normal correlation coefficient for both models indicate that the artificial neural network is better in prediction of total flavonoid yield than CCD. The optimal conditions using the desirability function at CCD model was achieved for the extraction time of 32.5 min, ethanol concentration of 100% (v/v) and solid-to-liquid ratio of 1:32.5 (m/v). The predicted yield at these conditions was 2.11 g/100 g of the dried extract (d.e.), while the experimentally obtained yield was 2.39 g/100 g d.e. The extraction process was optimized by the use of the simplex method for the MLP model. The optimal value of total flavonoid yield (2.80 g/100 g d.e.) was achieved after the extraction time of 27.2 min using ethanol concentration of 100% (v/v) at solid-to-liquid ratio of 1:20.7 (m/v). The predicted response value under optimal conditions for MLP model was also experimentally confirmed (2.71 g/100 g d.e.).

**Keywords:** artificial neural network, response surface methodology, extraction, total flavonoid, *Camellia sinensis*.

Available online at the Journal website: <http://www.ache.org.rs/HI/>

Green tea (*Camellia sinensis*) [1] can be found in the form of dried buds and leaves [2] on the market. Processing, packaging, storage and types of plants used are crucial for its quality. Also, the color, taste and quality of green tea depend on how the plant leaves are treated after harvest. Green tea has attracted the attention of people in the prevention of various diseases, ranging from weight loss to cancer [3]. The study of green tea carcinogenesis in animals confirmed its preventive effects on breast, prostate, lung and skin cancer [4]. The beneficial effects of green tea are attributed to its polyphenolic compounds. Tea catechins constitute about 80–90% of total flavonoids, while the content of flavonols is less than 10% [5]. Tea polyphenols, such as ketehin and flavonols remove the reactive oxygen species [6,7] and metal ions [6,8]. These flavonoids are antioxidants with phenolic hydroxyl groups [7]. The tea flavonoids remove NO and peroxynitrite,

produced from superoxide radicals and NO [9,10]. Certain polyphenols inhibit cell growth and proliferation, particularly in transformed cells, and also accelerate cell death. Tea polyphenols enhance the activity of antioxidant enzymes and hence antioxidant defense. Also, the polyphenolic components of green tea reduce LDL fat peroxidation of endothelial cells and therefore inhibit heme oxygenase gene expression [11]. These observations may be important, because the heme oxygenase is related to the transformation of mono-cyte-macrophages [12].

The determination of total flavonoids proposed in the literature was analyzed by the use of indirect VIS spectrophotometric methods based on the formation of complexes between the flavonoids and aluminum chloride [13–16]. Aluminum chloride builds the acid stable complexes with C-4 keto groups and/or C-3 or C-5 hydroxyl groups of flavones and flavonols. In addition, the aluminum chloride builds the acid unstable complexes with o-dihydroxyl groups A or B-ring flavonoids [17]. The flavones complexes with C-3 and C-5 hydroxyl groups, as well as o-dihydroxyl groups have an absorption maximum in the range of 415–440 nm.

Correspondence: I.M. Savić, Faculty of Technology, University of Niš, Bulevar oslobođenja 124, 16000 Leskovac, Serbia.

E-mail: [ici\\_teh@yahoo.com](mailto:ici_teh@yahoo.com)

Paper received: 13 March, 2012

Paper accepted: 15 May, 2012

SCIENTIFIC PAPER

UDC 66.061.3:663.951:004.94

*Hem. Ind.* 67 (2) 249–259 (2013)

doi: 10.2298/HEMIND120313066S

Response surface methodology (RSM) is a conventional method and effective statistic technique for optimization of complex processes. It has been successfully demonstrated that RSM can be used to optimize the isolation of total flavonoids from many medicine plants [18–22].

An artificial neural network (ANN) is a flexible mathematical structure which is capable of identifying the complex nonlinear relationships between input and output data sets. ANN models have been found useful and efficient, particularly in problems for which the characteristics of processes are difficult to describe using mathematical equations [23].

The optimization simplex method [24–27] is a very effective technique for finding the best process parameter values leading to optimal conditions. The simplex method for linear programming [28], developed by Dantzig [29], is perhaps the most well-studied algorithm in the optimization literature.

Moreover, personal computers, statistical software and computer graphics for desired function methodology implementation are now available and have been successfully applied in various processes and researches to optimize conditions for sample preparation and analysis of analytes. Carro and Lorenzo [30] used the desirability function to simultaneously optimize the solid-phase extraction of organochlorine and organophosphorus pesticide. Jimidar *et al.* [31] applied the desirability function the selection of optimum separation conditions in capillary zone electrophoresis. Bourguignon and Massart [32] simultaneously optimized several chromatographic performance goals using the desirability function.

Due to the health benefits of tea flavonoids, there is increasing interest their extraction. Since the optimization of total flavonoid extraction from green tea has not yet been described in the literature, the aim of this paper was to optimize this process using the ANN and RSM, as well as to compare these models. These approaches in optimization of extraction process have the advantage over other conventional techniques, owing to the small number of performed experiments. Extraction time, ethanol concentration and solid-to-liquid ratio (solvomodule) were selected as the independent variables, while the amount of total flavonoids was selected as the dependent variable.

## EXPERIMENTAL

*Samples and reagents.* Standard of rutin was purchased from Merck Chemicals Ltd. (United Kingdom). Absolute ethanol and ethanol of 96% (v/v) were purchased from Zorka Pharma (Serbia) and Alkaloid AD (Macedonia), respectively.

*Plant material.* Green tea (*Camellia sinensis*) was purchased from AD Aleva (Serbia) and dried to the

moisture content of 6% at room temperature in a dark place. The fine powder of tea was obtained by grounding the plant material in the electrical mill to the average particle size of 0.4 mm.

*Apparatus.* A double-beam Varian Cary-100 Conc. UV-Vis spectrophotometer, connected to a computer with Cary WinUV software, was used for measuring absorbance. The instrument has an automatic wavelength accuracy of 0.1 nm and matched quartz cells of the 10 mm cell path length.

*Extraction procedure.* Green tea powder (2 g) was transferred to a 100 cm<sup>3</sup> round bottom flask and covered with the selected volume of solvent. The ethanol concentration was varied according to CCD in order to investigate the effects on the yield of total flavonoid. All extractions were performed under reflux at the boiling temperature of the solvent. The flask was put in a thermostatic water bath. After the extraction, the solid matrix was separated from the liquid phase by filtering. The extract was evaporated under reduced pressure on a rotary evaporator at a temperature of 50 °C. After that, it was dried to constant mass in a desiccator and analyzed by the indirect UV-Vis method.

*Determination of total flavonoid.* The aluminum chloride colorimetric method was modified from the procedure reported by Woisky and Salatino [33]. Rutin was used to make the calibration curve. Rutin (10 mg) was dissolved in 80% (v/v) ethanol and then diluted in the range of 5–100 µg cm<sup>-3</sup>. The diluted standard solutions (0.5 cm<sup>3</sup>) were separately mixed with 1.5 cm<sup>3</sup> of 96% (v/v) ethanol, 0.1 cm<sup>3</sup> of 10% (m/v) aluminum chloride, 0.1 cm<sup>3</sup> of 1 mol dm<sup>-3</sup> potassium acetate and 2.8 cm<sup>3</sup> of distilled water. After incubation at room temperature for 30 min, the absorbance of reaction mixture was measured at the wavelength of 415 nm. Quartz cuvettes (1 cm×1 cm) were used for recording at room temperature. The amount of 10% (m/v) aluminum chloride was substituted by the same amount of distilled water in the blank. Similarly, 0.5 cm<sup>3</sup> of ethanol extracts were reacted with the aluminum chloride for determination of flavonoid content, as described above.

*Experimental design.* RSM consists of an adjustment of empirical models to the data obtained experimentally. Linear and quadratic mathematical models are employed to describe the system to be optimized [34]. If there are several factors that influence the particular system, it is necessary to use a screening design to investigate the significance of variables. The screening design, such as the factorial design 2<sup>f</sup>, can be used to meet this objective [14].

The first-order effects can be determined on the basis of factorial design, but if the second-order effects are observed, it is necessary to perform additional experimental runs. Thus, the second-order model is used

to evaluate the form of the true response. The number of additional experiments can vary due to the number of replicated center point. RSM used a three-factor and rotatable CCD consisting of 16 experimental runs, where two of the experiments refer to the center point of design. This point is commonly replicated in order to improve the precision of experiment. The set of  $2^3$  experiments present the cube points, while  $2 \times 3$  experiments are the axial points of design. The experimental runs were randomized to minimize the effects of unexpected variability in the observed responses. The variables were coded according to the following equation (Eq. 1):

$$x_i = (X_i - X_0) / \Delta X_i \quad (1)$$

where  $x_i$  is the coded value,  $X_i$  is corresponding actual value,  $X_0$  is the actual value in the center of the domain, and  $\Delta X_i$  is the increment of  $X_i$  corresponding to a variation of 1 unit of  $x$ .

The model of CCD is given by Eq.(2), where  $Y$  is the response;  $x_1$ ,  $x_2$  and  $x_3$  – the independent variables;  $a_0$  – the intercept of the  $y$  axis;  $a_1$ ,  $a_2$ ,  $a_3$ ,  $a_{11}$ ,  $a_{22}$ ,  $a_{33}$ ,  $a_{12}$ ,  $a_{13}$  and  $a_{23}$  – the various coefficients of the models (linear and quadratic):

$$y = a_0 + a_1x_1 + a_2x_2 + a_3x_3 + a_{11}x_1^2 + a_{22}x_2^2 + a_{33}x_3^2 + a_{12}x_1x_2 + a_{13}x_1x_3 + a_{23}x_2x_3 \quad (2)$$

The analysis of experimental design data and calculation of the predicted response were carried out by the use of Statistica software (version 8.0, StatSoft Inc., Tulsa, USA). After applying the fractional factorial design, the process parameters were identified for further optimization process using CCD. The results of that investigation are not presented in this study. The process variables and their levels are presented in Table 1.

The effect of extraction time, ethanol concentration and solvomodule at five variation levels is shown in Table 2. Thus, based on different combinations of the experimental conditions, it is possible to investigate the influence of these independent variables on the yield of total flavonoid, as well as the interactions between the process variables.

**Artificial neural network.** The experimental runs, used in CCD for modeling of the process, were also applied to describe the extraction of tea flavonoids using ANN. The ANNs were built using the same software as for the CCD model.

ANN is composed of a large number of neurons working in parallel providing an output response to an input data. All neurons are classified in three or more layers, depending on the number of layers in the hidden layer. In addition, there is one input and one output layer. Each neuron is linked to certain of its neighbours with varying coefficients of connectivity that represent the strengths (the weights) of these connections. The input layer receives input data, while the output predicts the value of selected response. The neurons in the hidden layer neither receive data nor predicts a response. The neuron sums the product of each connection weight ( $w_{kj}$ ) from a neuron  $j$  to the neuron  $k$  and input ( $x_j$ ) and the additional weight called the bias to get the value sum for the neuron  $k$  (Eq. (3)):

$$sum_k = \sum_{j=1}^p x_j \omega_{kj} + bias_k \quad (3)$$

Using an activation function, the sum of weighted inputs is transformed into output signal. Generally, the transfer functions are sigmoidal function, hyperbolic tangent and linear function, of which the most widely used for non-linear relationship is the sigmoidal function [35–43]. The general form of this function is given as follows (Eq. (4)):

$$y_j = f(x_i) = \frac{1}{1 + e^{-x_j}} \quad (4)$$

In addition, the exponential function is also used as an activation function in this case. The process of learning means the correction of weights to give the correct answers. Thus, the aim of training is to build a network, which will give correct prediction for the selected response. The MLP neural network architectures were trained in this study. MLPs are feed-forward neural networks trained with the standard backpropagation algorithm. Mathematically, this process of modeling can be described as in Figure 1. In this study, the train sample size was 70%, the test sample size was 15% and the validation sample size was 15%.

The learning period was completed when minimum RMS was reached (Eq. (5)):

$$RMS = \sqrt{\frac{\sum (\hat{y}_i - y_i)^2}{n}} \quad (5)$$

Table 1. Independent variables and their levels in the CCD

Independent variables	Symbol		Factor level				
	Uncoded	Coded	$-\alpha$	$-1$	$0$	$+1$	$+\alpha$
Extraction time, min	$\tau$	$X_1$	5	16.1	32.5	48.9	60
Concentration of ethanol, %	$C_e$	$X_2$	20	36.2	60	83.8	100
Solvomodule, m/v	$\omega$	$X_3$	10	16.1	25	33.9	40

Table 2. The experimental runs with the observed and predicted values of response for CCD and MLP model

Run	Sample	$\tau$ / min	$C_e$ / %	$\omega$ , m/v	$Y_{\text{observed}}$ , g/100 g d.e.	$Y_{\text{predicted}}$ , g/100 g d.e.	
						CCD	MLP
1	Train	48.9	36.2	16.1	1.58	1.73	1.47
2	Test	60.0	60.0	25.0	2.03	1.56	2.06
3	Train	16.1	83.8	33.9	1.22	1.48	1.21
4	Train	32.5	60.0	10.0	1.87	1.23	1.99
5	Validation	16.1	36.2	16.1	0.96	0.97	0.37
6	Train	32.5	60.0	25.0	1.81	1.95	1.59
7	Train	48.9	36.2	33.9	1.95	1.81	1.97
8	Train	32.5	60.0	40.0	1.43	1.50	1.45
9	Train	16.1	83.8	16.1	0.69	1.24	0.46
10	Train	32.5	100.0	25.0	2.81	1.87	2.69
11	Train	48.9	83.8	16.1	0.31	1.02	0.68
12	Train	48.9	83.8	33.9	1.60	2.00	1.51
13	Train	5.0	60.0	25.0	0.60	0.49	0.68
14	Validation	16.1	36.2	33.9	0.62	0.32	0.35
15	Test	32.5	60.0	25.0	1.99	1.95	1.59
16	Train	32.5	20.0	25.0	1.13	1.49	1.52

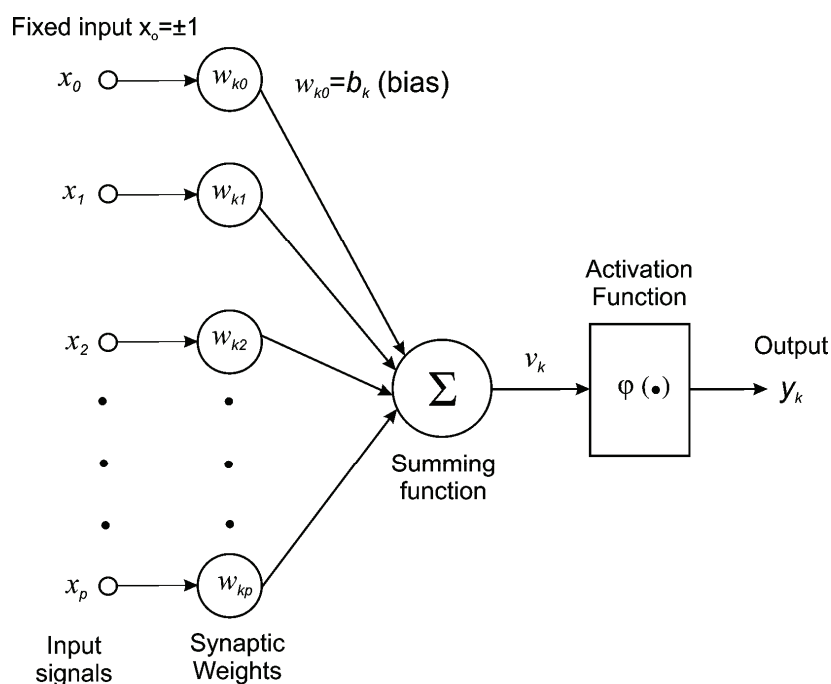


Figure 1. Schematic representation of ANN modeling.

where  $\hat{y}_i$  – the predicted values in the training or testing set,  $y_i$  – the observed values in the training or testing set,  $n$  – number of experimental runs.

Leave-one-out is the most commonly used form of cross-validation, where each data value is left out in turn and a model is derived using the remainder of the data. For every data point in the set, a value can be predicted and then compared with the true observed value. This is repeated for every data point in the set and permitted the calculation of a “cross-validated  $r^2$ ”

also written as  $Q^2$ . Cross validated  $r^2$  values are lower than normal ( $r^2$ ) and present a measure of quality of prediction. Unlike  $r^2$ , the cross-validated  $r^2$  can have negative values. The cross validated  $r^2$  can be calculated in the following way (Eq. (6)):

$$Q^2 = 1 - \frac{\sum_{i=1}^n (\hat{y}_i - y_i)^2}{\sum (y_i - \bar{y})^2} \quad (6)$$

where  $\hat{y}_i$  – the predicted values,  $y_i$  – the observed values,  $\bar{y}$  – the average observed values,  $n$  – number of experimental runs. A model is considered acceptable when the value of  $Q^2$  is higher than 0.5.

## RESULTS AND DISCUSSION

Extraction of the bioactive compounds from the medical plant materials has wide application for the phytopreparations production. Each procedure of phytopreparations production includes the extractions of bioactive compounds from the plant materials [44].

Extraction of the bioactive compounds from the plant material is the process of mass transfer in the solid–liquid system [45]. Factors such as the type of solvent, particle size, hydrodynamic conditions, temperature and density of plant material, affect the mass transfer and extraction degree. The selection of solvent type is critical for the yield and extraction degree, as well as the economy of the extraction process. The mass transfer rate is higher when the particle size decreases, due to the lower distance which the solvent has to pass through the solid phase. The hydrodynamic conditions, as well as the solid-to-liquid ratio have great effects on the extraction yield. Extraction can be performed at the solid-to-liquid ratio which provides a high extraction efficiency with less energy consumption and the use of simple devices. Increasing the temperature commonly improves the solubility of extracted compounds. Although, there are situations when the temperature has a negative impact due to poor solubility of bioactive compounds at higher temperatures.

Maceration is one-step extraction of the bioactive compounds from the herbal drugs at the defined temperature and solid-to-liquid ratio by the solvent with or without mixing. The commonly used solvents are volatile organic compounds and, for this reason, it is necessary to perform the maceration under reflux. An extract is separated from the plant material by filtering and by further pressing.

### Experimental design

An experimental design was applied to model and optimize the process extraction of total flavonoid from green tea. CCD was used as the best model for optimization. Statistica 8.0 provides a possibility to build different polynomial models by the use of different options in the software. There are four possibilities, where two of them refer to the linear modeling, and the remaining two on the second-order modeling. The second-order polynomial models have an advantages over the linear models, because they are better for predictions of the selected response. Thus, the differences between the observed and predicted values are smaller.

The final predicted process model in terms of coded factors for total flavonoid content is given below by Eq. (7):

$$Y = 1.95 + 0.32X_1 - 0.33X_1^2 + 0.11X_2 - 0.1X_2^2 + 0.08X_3 - 0.21X_3^2 - 0.24X_1X_2 + 0.18X_1X_3 + 0.22X_2X_3 \quad (7)$$

When the coded values of variables were replaced with the actual values, the empirical equation was obtained. This model is presented by Eq. (8):

$$Y = 1.95 + 0.64\tau - 0.66\tau^2 + 0.22C_e - 0.19C_e^2 + 0.16\omega - 0.41\omega^2 - 0.49\tau C_e + 0.37\tau\omega + 0.45C_e\omega \quad (8)$$

The significant factors of the equation are the linear and quadratic terms of extraction time, as well as the interaction between the extraction time and ethanol concentration. Unlike the quadratic term of extraction time and interaction extraction time/ethanol concentration, the linear term of extraction time has a positive effects on the yield of total flavonoid.

The characteristics of the CCD model are presented over the calculated RMS value of 0.4219,  $r^2$  value of 0.5779 and  $Q^2$  value of 0.5771.

### Artificial neural network

A feed-forward neural network (MLP) trained by a back-propagation algorithm was selected to develop the prediction model. The performances of ANNs were compared based on values of RMS training, RMS testing,  $Q^2$  and  $r^2$ .

The selected MLP model had three layers: the first layer had three units, the second layer had seven units, and the third layer had one output unit. In addition to these units, the architecture of this model contained two more bias in the input layer and in the hidden layer. The number of neurons in the hidden layer was selected after achieving a minimum error of the proposed model. The hyperbolic tangent function was used as an activation function in the hidden layer, while the logistic sigmoidal function was used in the output layer. When the learning period was over, the MLP model was tested with test data. For built MLP model, the calculated RMS training was 0.1929, and RMS testing was 0.2836. The value of  $Q^2$  was 0.9139, while the value of  $r^2$  was 0.9572.

The three-dimensional diagram of functional dependency between the extraction time and ethanol concentration on the yield of total flavonoid from the green tea is presented in Figure 2. The model of CCD indicates that the extraction time significantly affects the amount of total flavonoid in the extracts (Figure 2a). At lower ethanol concentrations, the flavonoids yield is increased up to the extraction time of 50 min, and then it remains constant. The functionality for the MLP model is more complex than for the CCD model

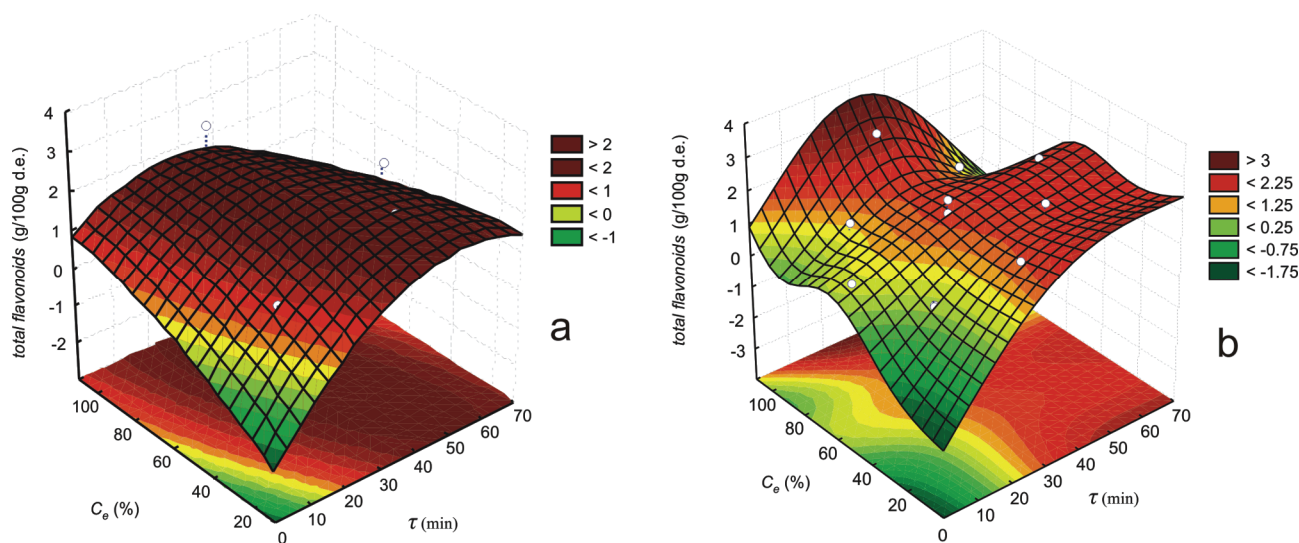


Figure 2. The functional dependency of extraction time and ethanol concentration on the yield of total flavonoid at the solvomodule of 1:25 (m/v) for: CCD (a); MLP (b).

(Figure 2b). Increasing the extraction time at lower ethanol concentrations leads to the increase of a response. At higher ethanol concentrations, the impact of extraction time has a positive effect on the flavonoid yield up to the extraction time of 35 min, and then it has a negative impact. By changing the ethanol concentration for short extraction times the influence on the yield is too small, while the yield increases for the extraction times from 30–40 min. It is very interesting that the yield decreases with increasing the ethanol concentration for longer extraction times.

The model of CCD for interaction between the extraction time and solid-to-liquid ratio is presented in Figure 3a. There is a strong interaction between these process parameters, which can be seen based on the function shape. The function maximum was noticed for

the extraction time from 30–40 and solvomodule from 1:25–1:40 (m/v). It can be seen from Figure 3b that the yield increases up to the extraction time of 35 min for lower solvomodules. At high solid-to-liquid ratios, the extraction time has a positive impact on the yield of total flavonoid. The significant effect of solvomodule was noticed for extraction times longer than 50 min. The flavonoid amount was increased by increasing the solvomodule.

The functional dependencies of ethanol concentration and solvomodule on the total flavonoid yield for both models are presented in Figure 4. The flavonoids amount decreased with increasing ethanol concentration at low solvomodules, while it decreased at high solvomodules. For lower ethanol concentrations, the solvomodule has a negative impact on the response.

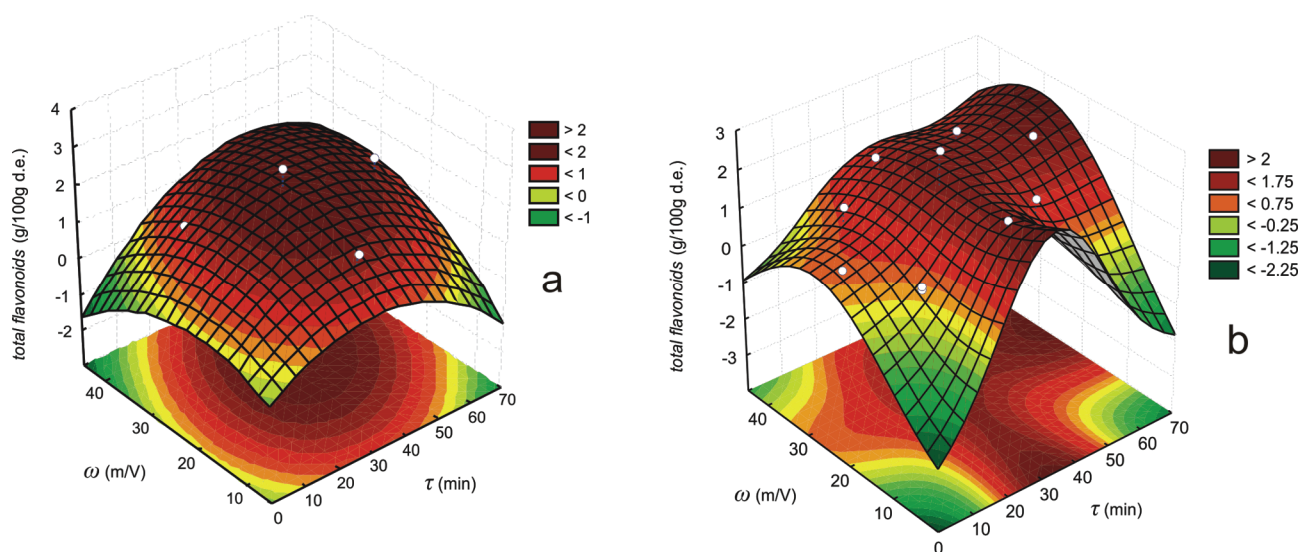


Figure 3. The functional dependency of extraction time and solvomodule on the yield of total flavonoid at the ethanol concentration of 60% (v/v) for: CCD (a); MLP (b).

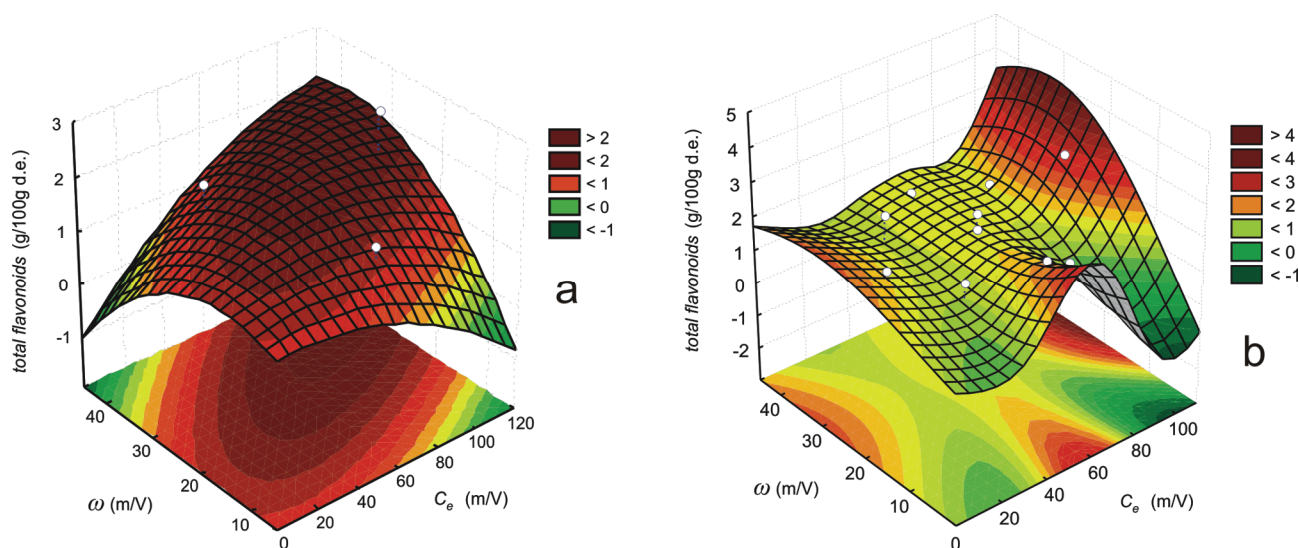


Figure 4. The functional dependency of ethanol concentration and solvomodule on the yield of total flavonoid at the extraction time of 32.5 min for: CCD (a); MLP (b).

Unlike the previous case, the solvomodule has a positive impact on the response for higher ethanol concentrations. The MLP model indicates that the yield decreases after the ethanol concentration of 60% (v/v) for lower solvomodules. As it can be seen from Figure 4b, the increase of ethanol concentration positively affects the flavonoids yield at solvomodules higher than 1:20 (m/v).

### The optimization of extraction process

RSM in combination with the desirability function approach has been proven to be a useful statistical tool to solve multi-variable problems and optimize one or several responses [46,47]. The desirability method makes use of an objective function,  $D(X)$ , called the desirability function and transforms an estimated response into a scale free value ( $d_i$ ) called desirability. The factor settings with maximum total desirability are considered to be the optimal parameter conditions. The simultaneous objective function is a geometric mean of all transformed responses (Eq. (9)):

$$D = (d_1 \times d_2 \times d_3 \times \dots \times d_n)^{1/n} = \left( \prod_{i=1}^n d_i \right)^{1/n} \quad (9)$$

where  $n$  is the number of responses in the measure. If any of the responses falls outside the desirability range, the overall function becomes zero.

Desirability is an objective function that ranges from zero outside of the limits, to one at the goal. The numerical optimization finds a point that maximizes the desirability function. Adjusting the weight or importance may alter the characteristics of a goal. For several responses, all goals get combined into one desirability function. For simultaneous optimization, each response must have a low and high value assigned to each goal.

The “Goal” field for responses must be one of five choices: “none”, “maximum”, “minimum”, “target” or “in range”. Factors will always be included in the optimization at their design range by default, or as a maximum, minimum of target goal.

The response optimization process was performed by selecting the software profile and desirability option. After applying the desirability function approach (Figure 5), the best optimized conditions were found to be the extraction time of 32.5 min, ethanol concentration of 100% (v/v) and solvomodule of 1:32.5 (m/v). The model of CCD predicts the value of 2.11 g/100 g d.e., while the experimentally obtained data is 2.39 g/100 g d.e.

For a general linear program, the simplex method performs an exponential number of pivots for several well-known pivot selection rules. No pivot selection rule of polynomial time complexity is known for the simplex method that guarantees a polynomial number of pivot iterations in the worst case.

The optimal conditions for MLP were reached after 50 iterations using the simplex method (Figure 6). The optimal total flavonoid yield of 2.80 g/100 g d.e. was predicted after 27.2 min of extraction and using the 100% (v/v) ethanol at the solvomodule of 1:20.7 (m/v). The validity of this predicted optimal value of total flavonoid was also experimentally confirmed (2.71 g/100 g d.e.), which is one more confirmation of the accuracy of the proposed MLP model.

### Comparing the optimization models

Based on the values of  $RMS$ ,  $Q^2$  and  $r^2$ , it was estimated which model is suitable for prediction of total flavonoid extraction from the green tea. The aim of modeling is to predict the response with minimum value of  $RMS$ , but with high values of  $Q^2$  and  $r^2$ . The cal-

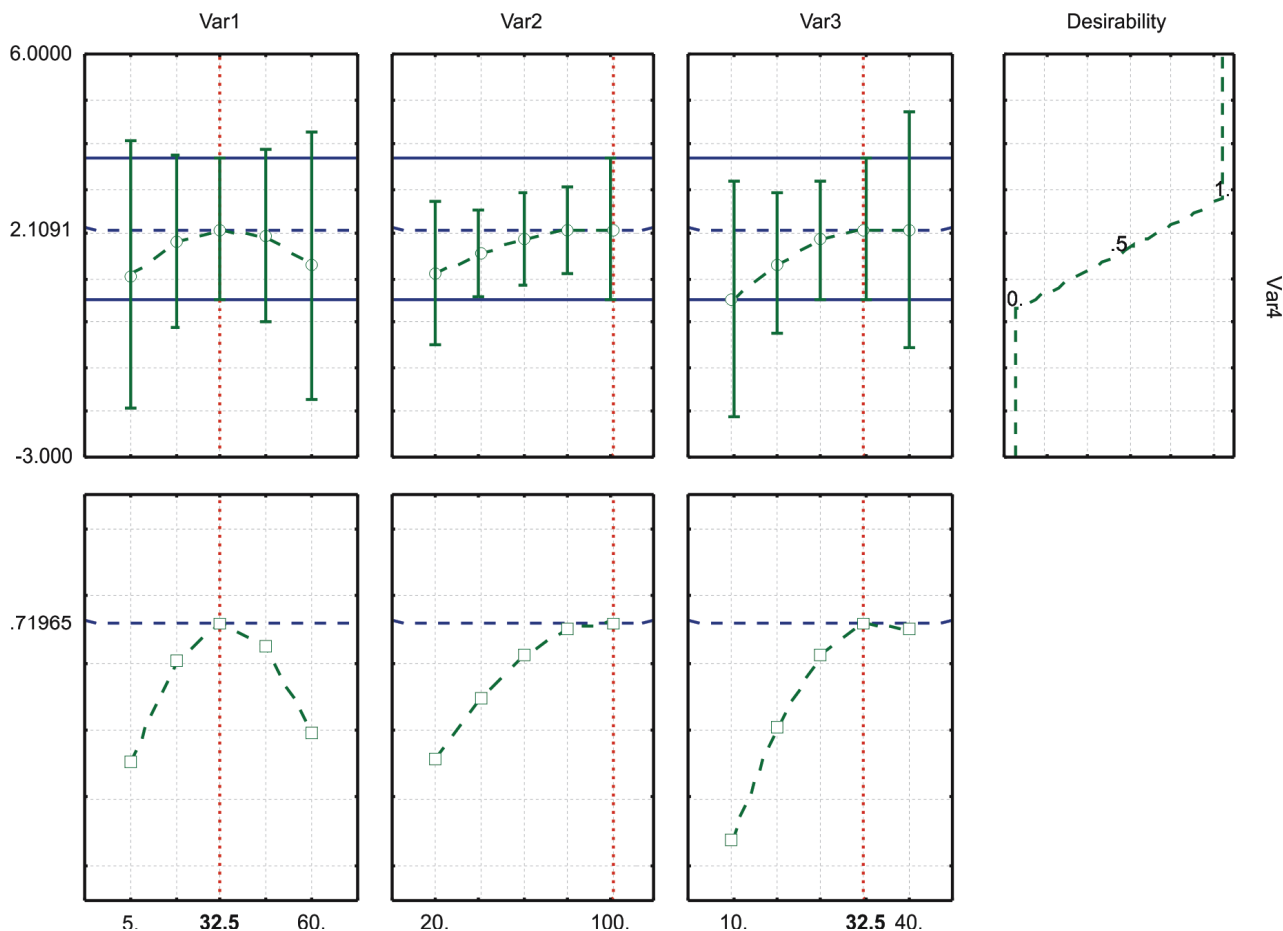


Figure 5. Desirability profile for optimization of process parameters.

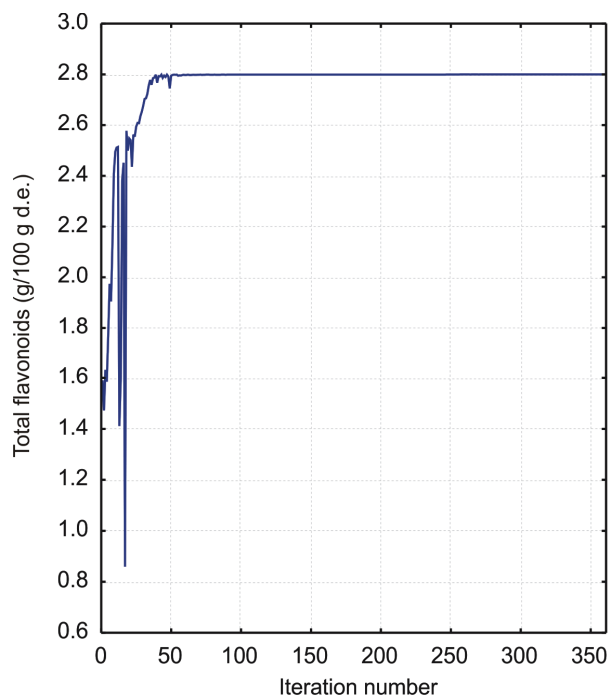


Figure 6. The optimization of the MLP model by simplex method.

culated values of *RMS* for the CCD model was 0.4219, and for the MLP model 0.1929. The values of  $Q^2$  and  $r^2$  for the CCD model were 0.9139 and 0.9572, while for the MLP model they were 0.5771 and 0.5779, respectively. The obtained values indicate that the MLP model has better performance for prediction of total flavonoid yield than the CCD model.

**CONCLUSION**

In this study, the extraction process of total flavonoid from green tea was modeled by the use of MLP and CCD models. The efficiency of both models was estimated in the aim of finding the best model for describing the extraction process. Based on the values of *RMS*,  $Q^2$  and  $r^2$ , the MLP model was proven as superior to the CCD model for prediction of the response. The optimal conditions of extraction process were obtained by using the desirability function for the CCD model and the simplex method for the MLP model. The agreement of experimental value of total flavonoid yield, obtained under optimal conditions, and the predicted value is a confirmation of the accuracy of the proposed models. The advantages of created mathematical models are the reduction of time and number of experi-



ments compared with the “one variable at a time” approach.

### Abbreviations

ANN – artificial neural network;  
ANOVA – analysis of variance;  
CCD – central composite design;  
LDL – low-density lipoprotein;  
MLP – multilayer perceptron;  
RMS – root mean square;  
RSM – response surface methodology.

### Acknowledgements

This work was supported by the Ministry of Education, Science and Technological Development of the Republic of Serbia under the project TR-34012. Ivan Savić is a recipient of a scholarship granted by the Ministry of Education, Science and Technological Development of the Republic of Serbia. The authors are thankful to reviewers for the useful comments and suggestions.

### REFERENCES

- [1] N.P. Seeram, S.M. Henning, Y. Niu, R. Lee, H.S. Scheuller, D. Heber, Catechin and caffeine content of green tea dietary supplements and correlation with antioxidant capacity, *J. Agr. Food Chem.* **54** (2006) 1599–1603.
- [2] H. Mukhtar, N. Ahmad, Tea polyphenols: Prevention of cancer and optimizing health, *Am. J. Clin. Nutr.* **71** (2000) 1698S–1702S.
- [3] N.T. Zaveri, Green tea and its polyphenolic catechins: Medicinal uses in cancer and noncancer applications, *Life Sciences* **78** (2006) 2073–2080.
- [4] H.N. Graham, Green tea composition, consumption, and polyphenol chemistry, *Prev. Med.* **21** (1992) 334–350.
- [5] D.C. Chu, Green tea-its cultivation, processing of the tea leaves for drinking materials, and kinds of green tea, In: L.R. Juneja, D.C. Chu, M. Kim, (Eds.), *Chemistry and Applications of Green Tea*, CRC Press, Boca Raton, FL, 1997.
- [6] M. Kim, M. Masuda, *Cancer Chemoprevention by Green Tea Polyphenols*, CRC Press, Boca Raton, FL, 1997.
- [7] C.S. Yang, P. Maliakal, X. Meng, Inhibition of carcinogenesis by tea, *Annu. Rev. Pharmac. Toxicol.* **42** (2002) 25–54.
- [8] V.E. Steele, D. Bagheri, D.A. Balentine, C.W. Boone, R. Mehta, M. A. Morse, S. Sharma, C.C. Sigman, G.D. Stoner, M.J. Wargovich, J.H. Weisburger, S. Zhu, G.J. Kelloff, Preclinical efficacy studies of green and black tea extracts, *Proc. Soc. Exp. Biol. Med.* **220** (1999) 210–212.
- [9] H.H. Chow, Y. Cai, D.S. Alberts, I. Hakim, R. Dorr, F. Shahi, J.A. Crowell, C.S. Yang, Y. Hara, Phase I pharmacokinetic study of tea polyphenols following single-dose administration of epigallocatechin gallate and polyphenon E, *Cancer Epidemiol. Biomarkers Prev.* **10** (2001) 53–58.
- [10] H.H. Chow, Y. Cai, I.A. Hakim, J.A. Crowell, F. Shahi, C.A. Brooks, R.T. Dorr, Y. Hara, D.S. Alberts, Pharmacokinetics and safety of green tea polyphenols after multiple-dose administration of epigallocatechin gallate and polyphenon E in healthy individuals, *Clin. Cancer Res.* **9** (2003) 3312–3319.
- [11] K.M. Pisters, R.A. Newman, B. Coldman, D.M. Shin, F.R. Khuri, W.K. Hong, B.S. Glisson, J.S. Lee, Phase I trial of oral green tea extract in adult patients with solid tumors, *J. Clin. Oncol.* **19** (2001) 1830–1838.
- [12] S.A. Laurie, V.A. Miller, S.C. Grant, M.G. Kris, K.K. Ng, Phase I study of green tea extract in patients with advanced lung cancer, *Cancer Chemoth. Pharm.* **55** (2005) 33–38.
- [13] C.C. Chang, M. Hua Yang, H.Mei Wen, J. Chuan Chern, Estimation of total flavonoid content in propolis by two complementary colorimetric methods, *J. Food Drug Anal.* **10** (2002) 178–182.
- [14] L. Alberto, L. Soares, V. Link Bassani, G. González Ortega, P. Ros Petrovick, Total flavonoid determination for the quality control of aqueous extractives from *Phyllanthus niruri* L., *Lat. Am. J. Pharm.* **22** (2003) 203–207.
- [15] D. Marinova, F. Ribarova, M. Atanassova, Total phenolics and total flavonoids in bulgarian fruits and vegetables, *J. Univer. Chem. Technol. Metall.* **40** (2005) 255–260.
- [16] E. Khatiwora, V.B. Adsul, M.M. Kulkarni, N.R. Deshpande, R.V. Kashalkar, Spectroscopic determination of total phenol and flavonoid contents of *Ipomoea carnea*, *Int. J. ChemTech Research* **2** (2010) 1698–1701.
- [17] T. Lundstedt, E. Seifert, L. Abramo, B. Thelin, A. Nyström, J. Pettersen, R. Bergman, Experimental design and optimization, *Chemom. Intell. Lab. Syst.* **42** (1998) 3–40.
- [18] Z. Sheng-tan, C. Guo-hua, L. Jing-ming, W. Tie-shan, Z. Ni, W. Zhao-yu, Optimization of *Jatropha curcas* for total flavonoids by response surface methodology and antibacterial activity, *J. Med. Plants Res.* **5** (2011) 3329–3334.
- [19] W. Liu, Y. Yu, R. Yang, C. Wan, B. Xu, S. Cao, Optimization of total flavonoid compound extraction from *Gynura medica* leaf using response surface methodology and chemical composition analysis, *Int. J. Mol. Sci.* **11** (2010) 4750–4763.
- [20] H.W. Wang, Y.Q. Liu, Y.H. Wang, Optimization of ultrasonic-assisted extraction of total flavonoids from leaves of the *Artocarpus heterophyllus* by response surface methodology, *Zhong Yao Cai* **34** (2011) 1125–1129.
- [21] W.H. Mao, L.J. Han, B. Shi, Optimization of microwave-assisted extraction of flavonoid from *Radix Astragali* using response surface methodology, *Sep. Sci. Technol.* **43** (2008) 671–681.
- [22] Z. Mei, P. Da-Ren, Z. Yi-Fei, A. Yu-Fang, Optimization of extraction process of flavonoids in *Rhododendron Pulchrum* sweet. based on artificial neural network combined with orthogonal experiment, *Journal of Xinyang Normal University (Natural Science Edition)* **2** (2011), doi: CNKI:SUN:XYSK.0.2011-02-032.

- [23] A.E. Giles, C. Aldrich, J.S.J. Van Deventer, Modeling of rare earth solvent extraction with artificial neural nets, *Hydrometallurgy* **43** (1996) 241–255.
- [24] C. Porte, Méthodes directes d'optimisation: méthodes d'optimisation à une variable et méthode Simplex, *Tech. Ing. P.* **228** (2002) 7–14.
- [25] F.H. Walters, S.L. Morgan, L.R. Paker, S.N. Deming, *Sequential Simplex Optimization*, CRC Press, Boca Raton, FL, USA, 1991.
- [26] S.N. Deming, S.L. Morgan, Simplex optimization of variables in analytical chemistry, *Anal. Chem.* **3** (1973) 278a–282a.
- [27] C. Porte, Méthodes directes d'optimisation: Méthodes dérivées de la method Simplex, *Tech. Ing. P.* **229** (2002) 1–8.
- [28] K.G. Murty, *Linear and Combinatorial Programming*, John Wiley & Sons Inc., New York, USA, 1976.
- [29] G.B. Dantzig, *Linear Programming and Extensions*, Princeton University Press, Princeton, NJ, 1963.
- [30] A.M. Carro, R.A. Lorenzo, Simultaneous optimization of the solid-phase extraction of organochlorine and organophosphorus pesticides using the desirability function, *Analyst* **126** (2001) 1005–1010.
- [31] M. Jimidar, B. Bourguignon, D.L. Massart, Application of Derringer's desirability function for the selection of optimum separation conditions in capillary zone electrophoresis, *J. Chromatogr., A* **740** (1996) 109–117.
- [32] B. Bourguignon, D.L. Massart, Simultaneous optimization of several chromatographic performance goals using Derringer's desirability function, *J. Chromatogr.* **586** (1991) 11–20.
- [33] R. Woisky, A. Salatino, Analysis of propolis: some parameters and procedures for chemical quality control, *J. Apicult. Res.* **37** (1998) 99–105.
- [34] R.F. Teófilo, M.M.C. Ferreira, Chemometrics II: spreadsheets for experimental design calculations, a tutorial, *Quim. Nova* **29** (2006) 338–350.
- [35] M. Dornier, M. Decloux, G. Trystram, A. Lebert, Dynamic modelling of crossflow microfiltration using neural networks, *J. Membrane Sci.* **98** (1995) 263–273.
- [36] H. Niemi, A. Bulsari, S. Palosaari, Simulation of membrane separation by neural networks, *J. Membrane Sci.* **102** (1995) 185–191.
- [37] E. Piron, E. Latrille, F. Rene, Application of artificial neural networks for crossflow microfiltration modelling: "Blackbox" and semi-physical approaches, *Comput. Chem. Eng.* **21** (1997) 1021–1030.
- [38] W.R. Bowen, M.J. Jones, H.N.S. Yousef, Dynamic ultrafiltration of proteins – A neural network approach, *J. Membrane Sci.* **146** (1998) 225–235.
- [39] W.R. Bowen, M.J. Jones, H.N.S. Yousef, Prediction of the rate of crossflow membrane ultrafiltration of colloids: A neural network approach, *Chem. Eng. Sci.* **53** (1998) 3793–3802.
- [40] N. Delgrange, C. Cabassud, M. Cabassud, L. Durand-Bourlier, J.M. Laine, Modelling of ultrafiltration fouling by neural networks, *Desalination* **118** (1998) 213–227.
- [41] M. Hamachi, M. Cabassud, A. Davin, M. Mietton Peuchot, Dynamic modelling of crossflow microfiltration of bentonite suspension using recurrent neural network, *Chem. Eng. Proces.* **38** (1999) 203–210.
- [42] C. Teodosiu, O. Pastravanu, M. Macoveanu, Neural network models for ultrafiltration and backwashing, *Water Res.* **34** (2000) 4371–4380.
- [43] S.M.A. Razavi, S.M. Mousavi, S.A. Mortazavi, Dynamic prediction of milk ultrafiltration performance, a neural network approach, *Chem. Eng. Sci.* **11** (2003) 4185–4195.
- [44] J. Asghari, B. Ondruschka, M. Mazaheritehrani, Extraction of bioactive chemical compounds from the medicinal Asian plants by microwave irradiation, *J. Med. Plants Res.* **5** (2011) 495–506.
- [45] V.D. Ponomarev, *Ekstragirovanie lekarstvennogo syr'ya*, Medicina, Moskva, 1976.
- [46] R.H. Myers, D.C. Montgomery, *Response Surface Methodology: Process and Product Optimization using Designed Experiments*, 2nd ed., John Wiley & Sons, Inc., New York, USA, 2002.
- [47] G. Derringer, R. Suich, Simultaneous optimization of several response variables, *J. Qual. Technol.* **12** (1980) 214–219.

## IZVOD

OPTIMIZACIJA EKSTRAKCIJE UKUPNIH FLAVONOIDA IZ *Camellia sinensis* PRIMENOM VEŠTAČKE NEURONSKE MREŽE I METODOLOGIJE POVRŠINSKOG ODGOVORAIvan M. Savić<sup>1</sup>, Vesna D. Nikolić<sup>1</sup>, Ivana M. Savić<sup>1</sup>, Ljubisa B. Nikolić<sup>1</sup>, Mihajlo Z. Stanković<sup>1</sup>, Karl Moder<sup>2</sup><sup>1</sup>Univerzitet u Nišu, Tehnološki fakultet, Leskovac, Srbija<sup>2</sup>University of Natural Resources and Life Sciences, Institute of Applied Statistics and Computing, Center of Experimental Design, Vienna, Austria

(Naučni rad)

Poznato je da slobodni radikali deluju vrlo nepovoljno na čovekov organizam, oštećujući ćelije tkiva i organa dovodeći do degenerativnih oboljenja i raznih oblika raka. Utvrđeno je da su polifenoli nekih biljaka jaki antioksidansi, a za zeleni čaj potvrđeno je da ima najveću količinu polifenola (300–400 mg). Zbog visoke koncentracije polifenola i flavonoida (5–27%), zeleni čaj se koristi kod infekcija, prehlada smanjujući oštećenja ćelija pod dejstvom virusa. Upotrebom zelenog čaja sprečava se razvoj malignih ćelija, a redovno konzumiranje je povezano sa smanjenom incidencom razvoja raka jajnika, jednjaka, mokraćne bešike, pluća, pankreasa, prostate i kože. Takođe, sprečava razvoj ateroskleroze i srčanih oboljenja, ubrzava metabolizam i sagorevanje masti. Snižava nivo holesterola, pa se primenjuje i kod povećanog nivoa holesterola u krvi. Cilj ovog rada bio je modelovanje i optimizacija procesa ekstrakcije ukupnih flavonoida iz zelenog čaja (*Camellia sinensis*) primenom veštačke neuronske mreže (eng. *artificial neural network*, ANN) i metodologije površine odgovora (eng. *response surface methodology*, RSM), kao i poređenje ovih optimizacionih tehnika. Vreme ekstrakcije, koncentracija etanola i solvomoduul posmatrani su kao nezavisno promenljive, dok je prinos ukupnih flavonoida odabran kao zavisno promenljiva. Centralni kompozitni dizajn (eng. *central composite design*, CCD), primenom polinomnog modela drugog reda i višeslojni perceptron korišćeni su za fitovanje dobijenih eksperimentalnih podataka. Vrednosti  $RMS$ ,  $Q^2$  i  $r^2$  za optimizacione modele ukazuju da je model „višeslojnog filtriranja“ (eng. *multilayer perceptron*, MLP) bolji za predviđanje prinosa ukupnih flavonoida od CCD modela. Kod CCD modela, primenom „poželjne“ (eng. *desirability*) funkcije, optimalni uslovi ekstrakcije ukupnih flavonoida postignuti su pri vremenu ekstrakcije od 32,5 min, koncentraciji etanola od 100% (v/v) i solvomoduulu od 1:32,5 (m/v). Pri datim uslovima, predviđeni prinos ukupnih flavonoida je 2,11 g/100 g suvog ostatka (s.o.), dok je eksperimentalno dobijena vrednost od 2,39 g/100 g s.o. Proces ekstrakcije optimizovan je primenom simpleks algoritma kod MLP modela. Po ovom modelu optimalni uslovi ekstrakcije ukupnih flavonoida iz zelenog čaja postizu se za vreme ekstrakcije od 27,2 min, pri koncentraciji etanola od 100% (v/v) i solvomoduulu od 1:20,7 (m/v). Pri ovim optimalnim uslovima ekstrakcije, predviđena vrednost prinosa ukupnih flavonoida (2,80 g/100 g s.o.) potvrđena je eksperimentalno (2,71 g/100 g s.o.).

*Ključne reči:* Veštačka neuronska mreža • Metodologija površine odgovora • Ekstrakcija • Ukupni flavonoidi • *Camellia sinensis*